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An Electrochemical Sensor Based on Novel Ion Imprinted Polymeric Nanoparticles for Selective Detection of Lead Ions

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In this study, the novel surface ion-imprinted polymer (IIP) particles were prepared and applied as an electrode modifier in stripping voltammetric detection of lead(II) ion. A carbon paste electrode (CPE) modified with IIP nanoparticles and multi-walled carbon nanotubes (MWCNTs) were used for accumulation of toxic lead ions. Various factors that govern on electrochemical signals including carbon paste composition, pH of the preconcentration solution, supporting electrolyte, stirring time, reduction potential and time were studied in details. The best electrochemical response for Pb(II) ions was obtained with a paste composition of 7% (w/w) of lead IIP, 10% MWCNTs, 53% (w/w) of graphite powder and 30% (w/w) of paraffin oil using a solution of 0.1 M acetat buffer solution (pH = 4.5) with a extraction time of 15 min. A sensitive response for Pb(II) ions in the concentration range of 3-55 $\mu\text{g l}^{-1}$ was achieved. The proposed electrochemical sensor showed low detection limit (0.5 $\mu\text{g l}^{-1}$), remarkable selectivity and good reproducibility (RSD = 3.1%). Determination of lead(II) content in different environmental water samples was also realized adopting graphite furnace atomic absorptions spectrometry (GF-AAS), and the obtained results were satisfactory.

Keywords: Lead, Imprinted polymer, Heavy metal, Electrochemical sensing, Carbon past electrode

INTRODUCTION

Presently, pollution of the environment by heavy metal ions (HMIs) has created a serious environmental problem [1]. Among all of the HMIs, lead is one of most common toxic bivalent metal ion because of its serious toxicity even at low concentrations [2]. The permissible limit of lead in drinking waters considered by the Environmental Protection Agency (EPA) is 15 $\mu\text{g l}^{-1}$ [3]. Serious poisoning effects of lead have led to remarkable efforts towards developing of sensitive analytical methods for the determination of lead [4]. Electrochemical methods have been attractive for detection of lead due to its experimental simplicity, low cost, remarkable sensitivity and portability [5,6]. Insofar, as selectivity is concerned, introducing of high selective

modifiers for electrochemical monitoring of lead(II) ion is very important. Hence, ion imprinting technology based on the highly selective ion-imprinted polymers (IIPs) can be considered as a most versatile and promising option to incorporate selective recognition sites in a polymeric matrix [7]. Traditionally, ion-imprinted polymers (IIPs) have been prepared by bulk or precipitation polymerization method [8]. However, the obtained IIPs suffer some common limitations such as low binding capacity, poor site accessibility for target species, irregular shape, slow mass transfer, and heterogeneous binding site distribution [9]. To avoid the disadvantages mentioned above, the surface ion imprinted technique has been developed due to some outstanding advantages such as low mass-transfer resistance, convenient to prepare, good accessibility to the target ions, higher affinity and selectivity [10]. Up to now, different surface imprinting materials based on a variety of supports such as multi-walled carbon nanotubes

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(MWCNTs) [11], magnetic nano-particle [12], reduced graphene oxide (RGO) [9], silica particles [13] and nanometer titanium dioxide (TiO_2) [14] have been exploited.

Recently, IIPs have been utilized as the selective modifiers for electrochemical measurement of some HMIs including mercury(II) [9], silver(I) [15], zinc(II) [16], palladium(II) [17], and copper(II) [18] ions. However, it is of great significance to develop electrochemical sensors based on nanosized IIP beads for detection of toxic HMIs in low concentrations due to better properties in term of selectivity and sensitivity [19]. The aim of the present work is to design a novel lead(II) electrochemical sensor based on the nano-structured IIP material and MWCNTs. Various experimental factors, linear calibration response, and the limit of detection (LOD) of the suggested electrochemical device in the detection of lead(II) were evaluated. The desired method was successfully employed for voltammetric detection of Pb(II) ions in various environmental water samples with satisfactory selectivity.

EXPERIMENTAL

Reagents and Apparatus

Ethylenglycol dimethacrylate (EGDMA), 2-vinylpyridine (2-VP), 2,2'-azobisisobutyronitrile (AIBN), 3-methacryloxypropyl trimethoxysilane (KH-570), ferrous sulfate heptahydrate ($\text{FeSO}_4 \cdot 7\text{H}_2\text{O}$), ferric chloride hexahydrate ($\text{FeCl}_3 \cdot 6\text{H}_2\text{O}$) and ammonium hydroxide (NH_4OH , 25%, w/w) were purchased from Merck (Darmstadt, Germany). The magnetite (Fe_3O_4) nanoparticles were prepared through coprecipitation of Fe^{2+} and Fe^{3+} ions in the presence of ammonium hydroxide solution according to the literature methods [20]. Graphite powder (average particle size 50 μm) and high viscosity paraffin oil (density = 0.88 Kg l^{-1}), both from Merck (Darmstadt, Germany), were used as the working electrode substrates.

All electrochemical data were obtained with a Metrohm computerized electroanalyzer Model 757 VA (Herisau, Switzerland). The electrochemical cell consisted of a home-made modified carbon paste electrodes (MCPE) as a working electrode and an Ag/AgCl electrode and a platinum wire as the reference and counter electrode, respectively.

The pH of buffer solutions was determined with a digital Metrohm 827 pH-meter (Herisau, Switzerland).

Synthesis of the Lead(II) Imprinted Polymeric Nanoparticles

Lead imprinted polymer based on surface imprinting process was prepared by the following procedure. Firstly, vinyl functionalization of the magnetic nanoparticles was performed by reaction of coupling agent KH-570 with Fe_3O_4 nanoparticles. For this purpose, 1.0 g Fe_3O_4 was suspended in 50 ml of acetonitrile followed by the addition of 1.1 ml of KH-570 and the mixture was refluxed at 60 °C for 24 h under stirring. The resulting particles were separated from the solution by a magnet and stored for further steps. As the next step, 0.5 mmol of $\text{Pb}(\text{NO}_3)_2$ and 2 mmol of 2-VP (monomer) were dissolved in 50 mL of DMSO-AN (1:2). After 2 h of the complexation reaction, 0.5 g of vinyl functionalized Fe_3O_4 was dispersed in the above solution for 5 min by ultrasonic dispersion. Afterwards, 10 mmol of EGDMA and 80 mg AIBN were added to the reaction vessel and the solution was mixed for a few minutes. Then, the oxygen of the sample solution was removed by bubbling nitrogen through the sample for 15 min. The mixture solution was heated at 60 °C in a water bath under N_2 atmosphere for 24 h. Afterward, the resultant product were washed successively with ethanol to remove the unreacted materials. The entrapped lead ions were then removed with a solution containing 0.5 M of HCl. Finally, the prepared IIP materials were rinsed with ultra-pure water seven times and dried at 60 °C for the further use. Schematic illustration for the synthesized IIP nanoparticles is represented in Fig. 1. Correspondingly, the non-imprinted polymer (NIP) was prepared in the same method but in the absence of the lead(II) ions.

Fabrication of MCPE

The MCPE was prepared by thoroughly mixing of 106 mg graphite powder, 60 μl paraffin oil, 14 mg of lead IIP and 20 mg of MWCNTs using a mortar and pestle. Then, the homogenized paste was packed into a Teflon tube (2 mm inner diameter and 10 cm long) and connected to a copper wire to provide the electrical connection. The fresh surface of the prepared electrode was obtained by replacing the carbon paste and polishing onto the weighing paper.

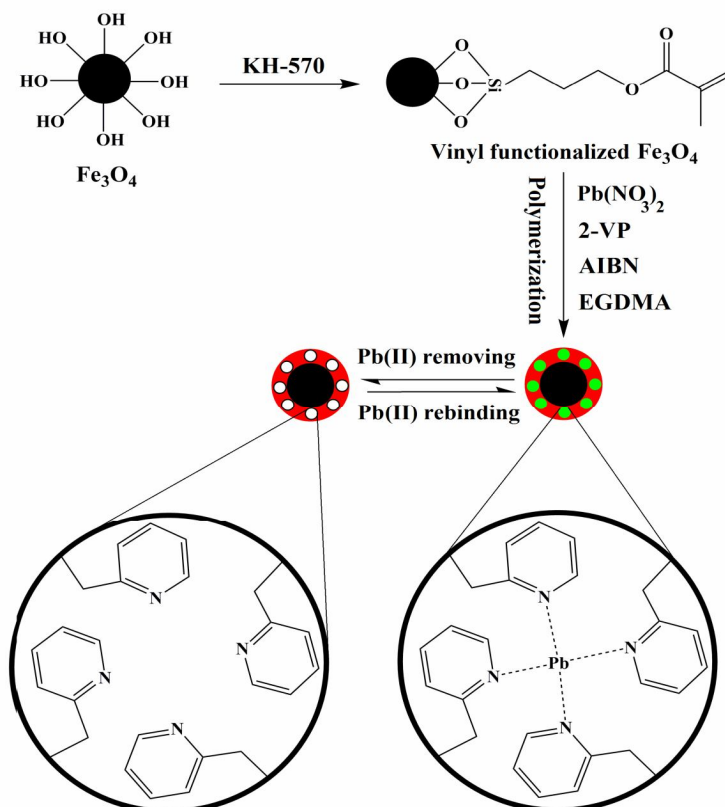


Fig. 1. Schematic depiction of the imprinting process for the preparation of lead IIP nanobeads.

Preparation of Water Samples

Various real water samples including river water (Kardeh river, Mashhad, Iran), aqueduct water (Balvard village, Kerman, Iran), waste water (copper factory, Sarchashmeh, Kerman, Iran) and waste water (Coal Processing, Zarand, Kerman, Iran) were selected to explore the applicability of the proposed method. Water samples were filtered through a membrane of 0.2 μm pore-size to remove any suspended material and stored at 4 $^\circ\text{C}$ in a refrigerator until analysis.

Electrochemical Procedure

The MCPE was immersed in a 20 ml of 0.1 M acetate buffer solution (pH = 4.5) containing a known amount of lead ions and the solution was stirred (500 rpm) for 15 min in an open circuit condition. Then, the MCPE was inserted into the washing solution (deionized water) for 15 s, and then placed into the voltammetric cell containing 20 ml of HCl solution (0.1 M) as supporting electrolyte. Finally,

preconcentrated $\text{Pb}(\text{II})$ was reduced for 40 s in -1.0 V. The differential pulse voltammograms were recorded between -0.8 to 0.0 V with pulse amplitude of 0.1 V and scan rate of 50 mV s^{-1} at room temperature (23 ± 2 $^\circ\text{C}$).

RESULTS AND DISCUSSION

Characterization

The Fourier transform infrared (FT-IR) spectra for Fe_3O_4 , vinyl functionalized Fe_3O_4 and lead IIP particles are demonstrated in Fig. 2. The adsorption band around at 629, 560 and 440 cm^{-1} are the characteristic bands of Fe_3O_4 , corresponding to Fe-O vibrations of Fe_3O_4 [21]. In vinyl functionalized Fe_3O_4 spectrum, the characteristic band at 816 cm^{-1} is assigned to Si-O bending and stretching vibrations [22]. The absorption peak around 1089 cm^{-1} was confidently attributed to the siloxane vibrations [23]. The band at 2925 cm^{-1} was assigned to the stretching vibrations of C-H bonds [24]. The observed peak at 1634 cm^{-1} can be

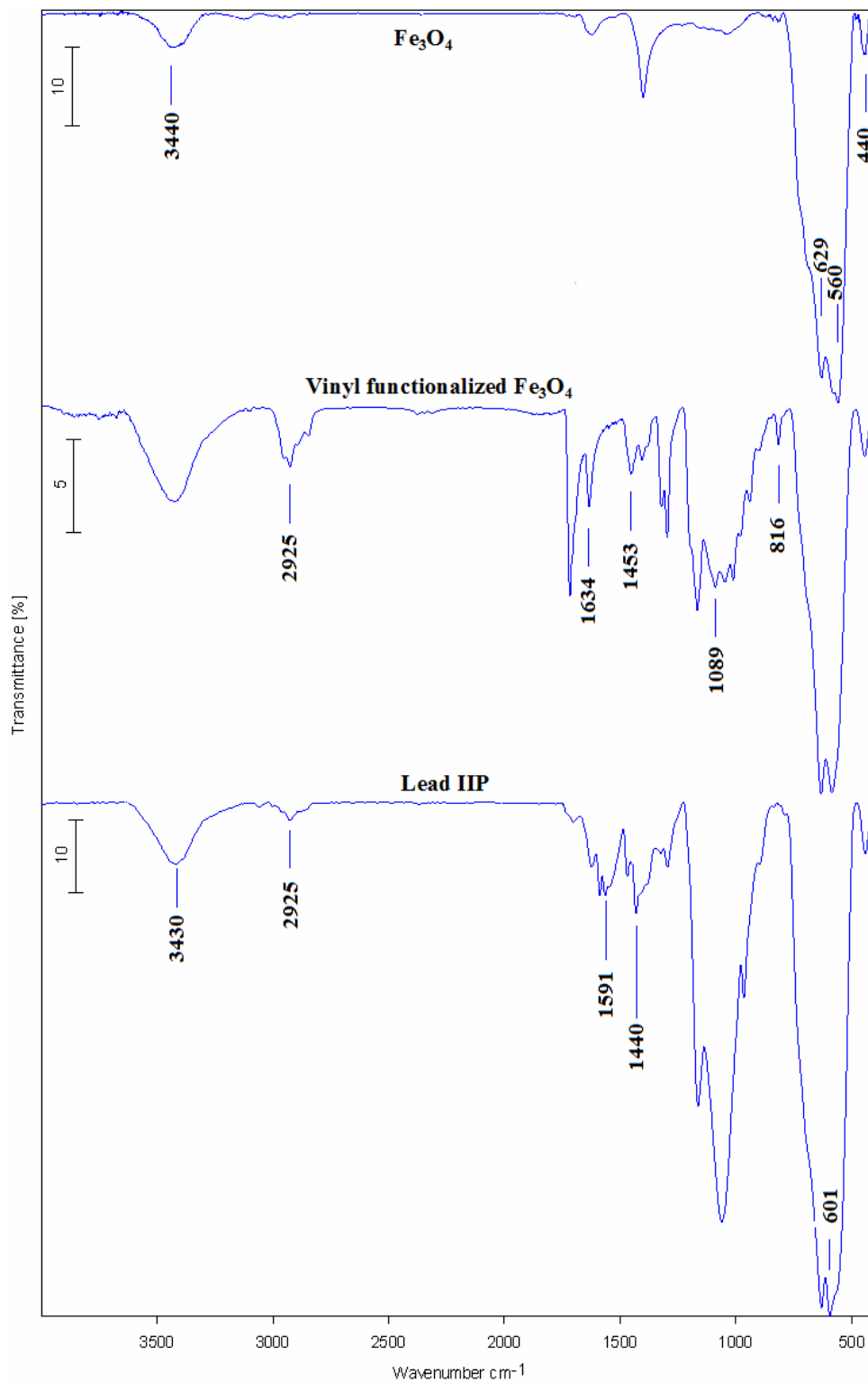


Fig. 2. FT-IR spectra of Fe₃O₄, vinyl functionalized Fe₃O₄ and lead IIP nanoparticles.

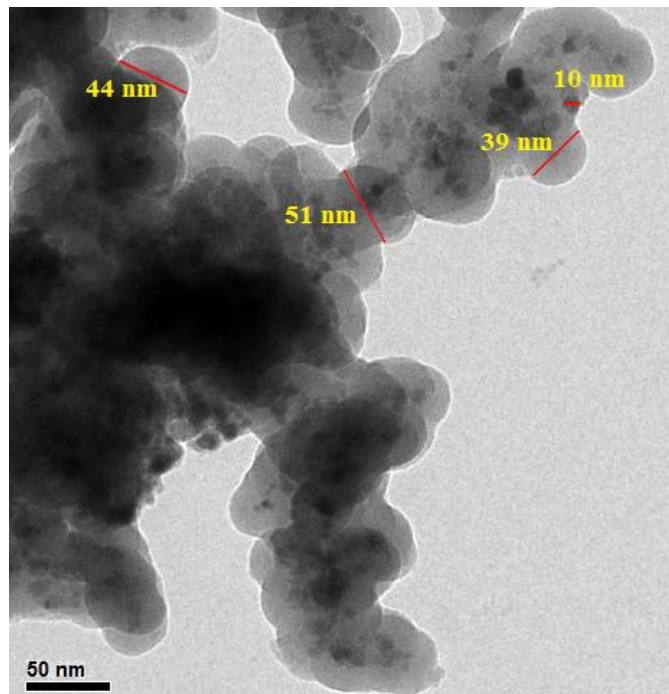


Fig. 3. TEM image of as-prepared IIP nanoparticles.

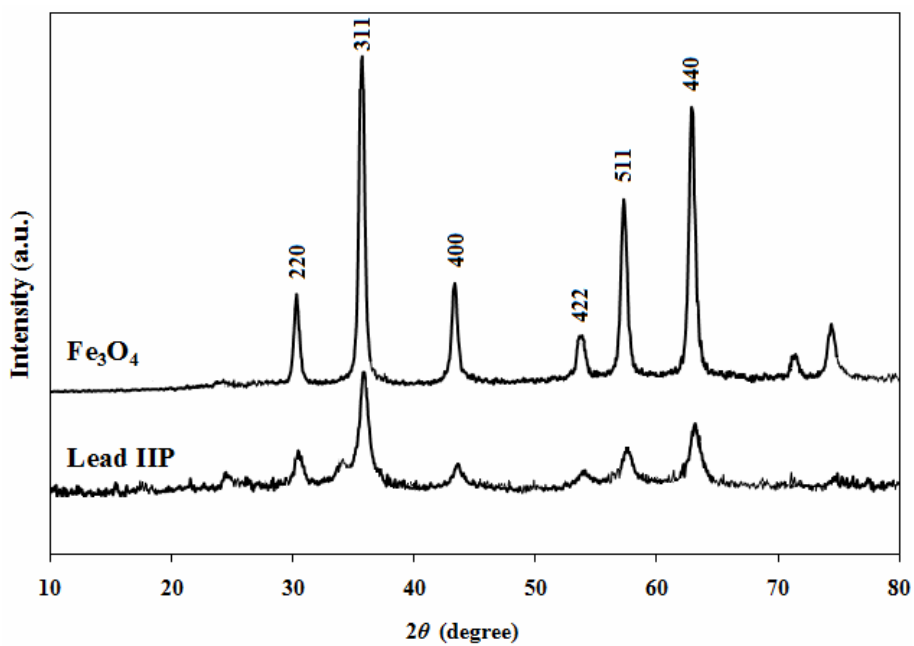


Fig. 4. XRD patterns of Fe₃O₄ and the lead IIP nanoparticles.

attributed to the C=C stretching band [25]. In FT-IR spectra of lead IIP particles, the bands at 3430, 2925, 1591 and 1601 cm^{-1} are assigned to O-H, C-H aliphatic, C=N and Fe-O stretching vibrations, respectively. Moreover, the -CH₂-CH₂- bending vibration absorption band was shown at 1450 cm^{-1} [26].

Transmission electron microscopy (TEM) image of the imprinted material is shown in Fig. 3. As can be seen, the surface of Fe₃O₄ nanoparticles was fully coated by polymer layer. So, it can be confirmed from TEM image that lead IIP nanoparticles have been successfully synthesized, and the core-shell structure formed. Moreover, the thickness of the imprinted layer was about 15-25 nm.

Figure 4 shows X-ray diffraction (XRD) patterns of Fe₃O₄ and the lead IIP samples. The XRD pattern of Fe₃O₄ nanoparticles shows six characteristic peaks that the diffraction peaks ($2\theta = 30.2^\circ, 35.7^\circ, 43.4^\circ, 53.8^\circ, 57.4^\circ$ and 63.1°) indexed to the (220), (311), (400), (422), (511), and (440) crystalline plane of Fe₃O₄ [27]. Furthermore, the decrease in the intensity of the characteristic peaks in lead IIP sample can be attributed to lower crystallinity of the Fe₃O₄ structure after modification with imprinted layer.

Electrochemical Behavior of Pb(II) at the MCPE

The electrochemical behavior of the MCPE for preconcentration of lead(I) ions were investigated by differential pulse voltammetry. Figure 5 shows the typical voltammograms of the unmodified CPE, MWCNTs-CPE, IIP-CPE, IIP-MWCNTs-CPE, NIP-CPE with lead(II) and IIP-MWCNTs-CPE (proposed MCPE) without lead(II) ions. The distinct voltammetric peaks indicate the ability of the various modifiers for sensing of analyte ion on the modified electrode surface. As seen in Fig. 5, the obtained response for the IIP-MWCNTs-CPE is higher noticeably than other modifiers, indicating properly preconcentration of lead(I) by MCPE.

Optimization of Significant Parameters

Considering the carbon paste composition is a critical parameter in the electrochemical response of the MCPE, the effect of the lead IIP amount on the voltammetric signal of 25 $\mu\text{g l}^{-1}$ of Pb(II) was studied. The results are shown in Fig. 6. The signal of Pb(II) increased with the increase of lead IIP amount to 7% (mass/mass). The peak current decreased

at higher IIP content because of the diminished conductivity of the carbon paste. Moreover, the best electrochemical signal with higher sensitivity achieved with 10% for MWCNTs in carbon paste composition. Consequently, 7% lead IIP, 10% MWCNTs, 53% graphite powder and 30% paraffin oil were selected as the optimal values in further experiments.

In order to evaluate influence of pH of the buffer solution on the peak current, the MCPE was applied for preconcentration of lead ions (25 $\mu\text{g l}^{-1}$) in pH range 3.5-5.5 under stirring condition for 10 min. As clearly seen in Fig. 7, an optimum pH of 4.5 was selected in next experiments.

The influence of stirring time (in the range of 5-20 min) on the proposed method was also investigated. For this purpose, the MCPE was applied for extraction of 25 $\mu\text{g l}^{-1}$ lead(II) solution (pH 4.5) at different times, while keeping all the other experimental parameters constant. As depicted in Fig. 8, the electrochemical signal increased linearly with changing extraction time from 1 to 12.5 min and remained almost constant at higher stirring times (>15 min) due to the surface saturation of the MCPE. Thus, a time of 15 min was employed as the stirring time in further studies.

The effect of the supporting electrolyte was evaluated by monitoring the differential pulse voltammograms in different media (all 0.1 M) including hydrochloric acid, phosphoric acid, nitric acid, and acetic acid. As the result, the highest signal with ideal peak shape and lowest background current was obtained in HCl medium. Moreover, the influence of HCl concentration on the analytical response of proposed sensor was also examined in the range from 0.001-1 M. It was found out that the peak current values increases with increasing the electrolyte concentration up to 0.1 M. However, the analytical signals were reduced when the acid concentrations were more than 0.1 M. Therefore, 0.1 M HCl was chosen in this work.

The dependence of reduction potential on the analytical signal intensity was studied by varying the reduction potential from -0.7 to -1.3 V vs. Ag/AgCl. When the reduction potential was more than -1.0 V, the peak current reduced, because of the cohydrogen evolution at such negative potentials. Taking into account that the electrochemical signals indicated well-defined peaks with maximum currents at potential of -1.0 V, this reduction potential was selected for the subsequent experiments.

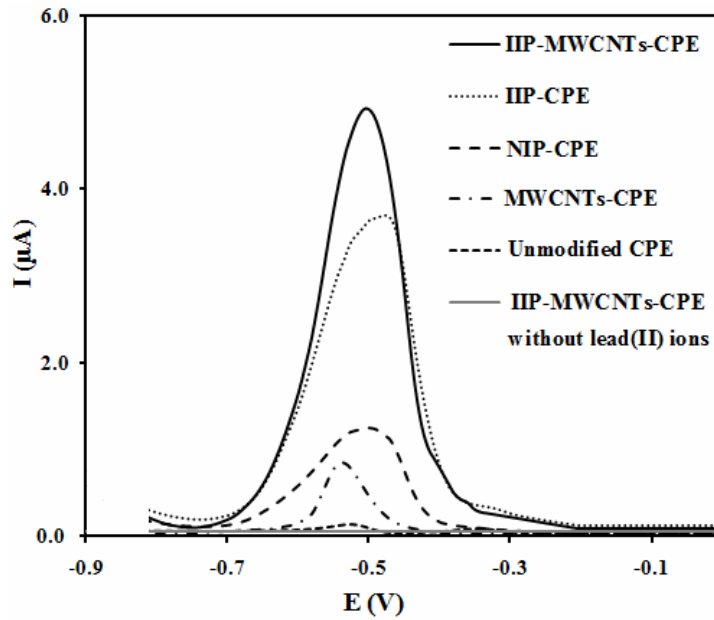


Fig. 5. Differential pulse voltammograms of unmodified CPE, MWCNTs-CPE, IIP-CPE, IIP-MWCNTs-CPE, NIP-CPE with $25 \mu\text{g l}^{-1}$ of Pb(II) and IIP-MWCNTs-CPE without Pb(II) in extraction medium. Other conditions; stripping solution = 0.1 M potassium nitrate and acetate buffer (0.1 M, pH 4.0), extraction time = 10 min, amount of IIP = 5%, accumulation potential = -0.9 V (vs. Ag/AgCl), accumulation time = 40 s, scan starting from -0.8 to 0.0 V, scan rate = 50 mV s^{-1} , pulse amplitude = 0.1 V and pulse period = 4 ms.

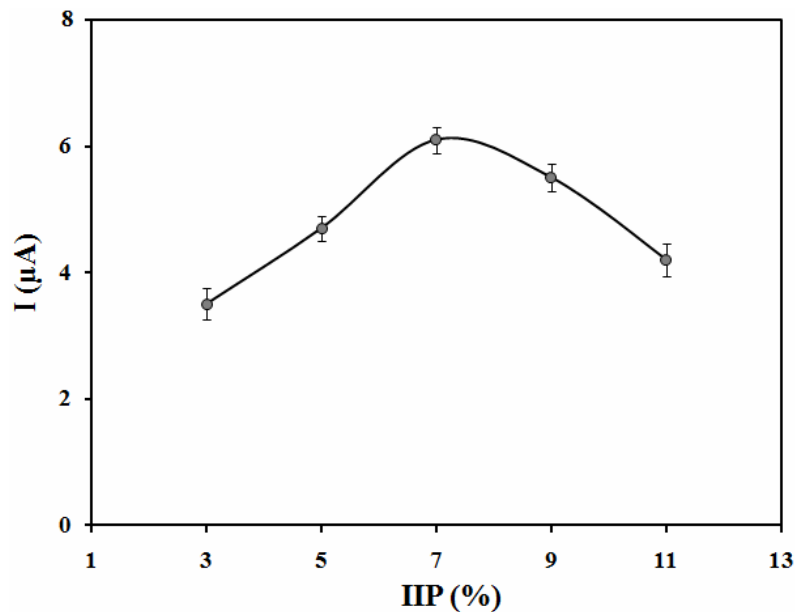


Fig. 6. The effect of lead IIP amount on the electrode response (MWCNTs = 10%).

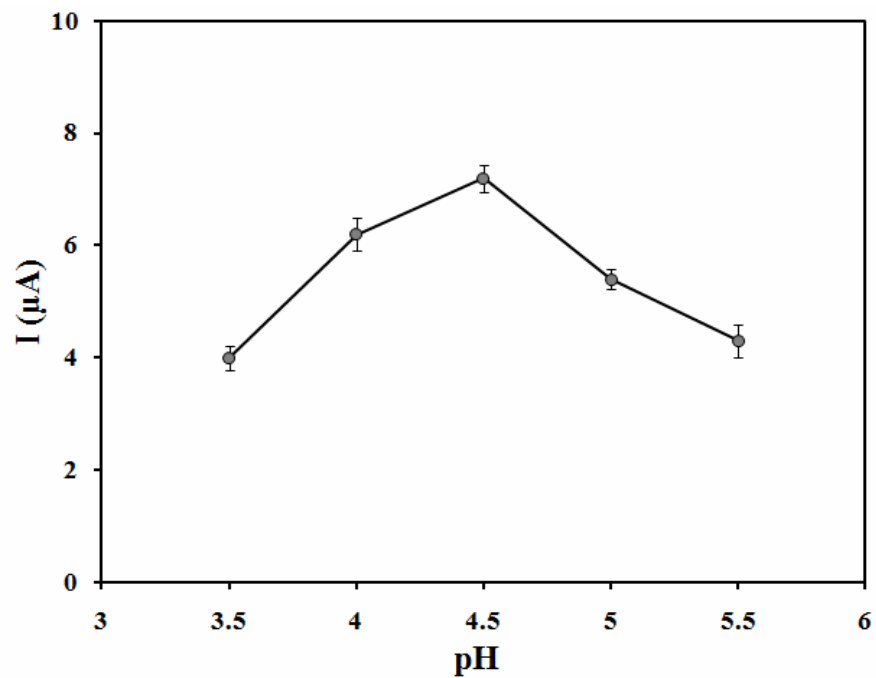


Fig. 7. Effect of preconcentration solution pH on the voltammetric signal.

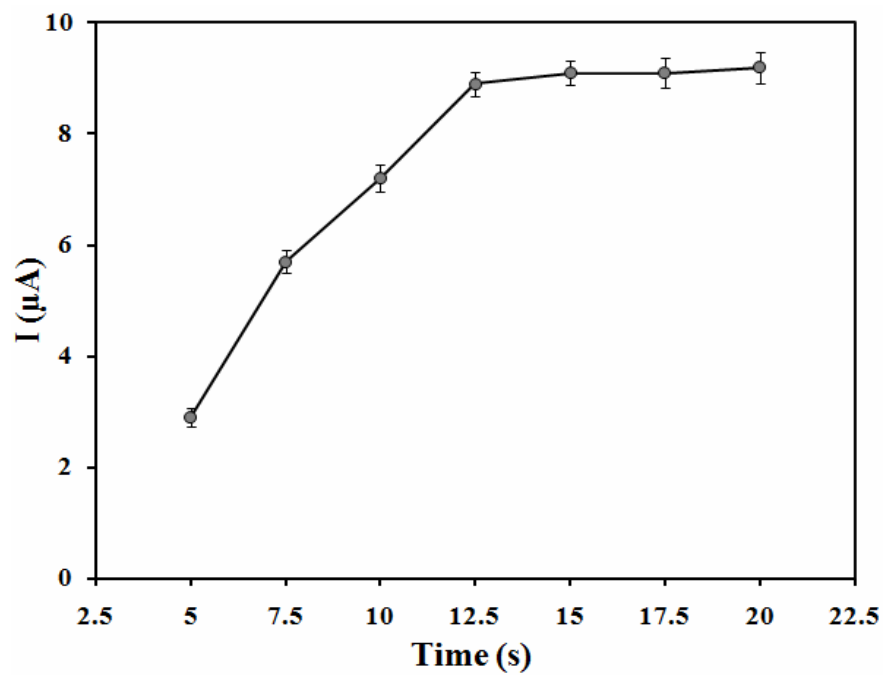


Fig. 8. Peak current response versus the extraction time for $25 \mu\text{g l}^{-1}$ of Pb(II) .

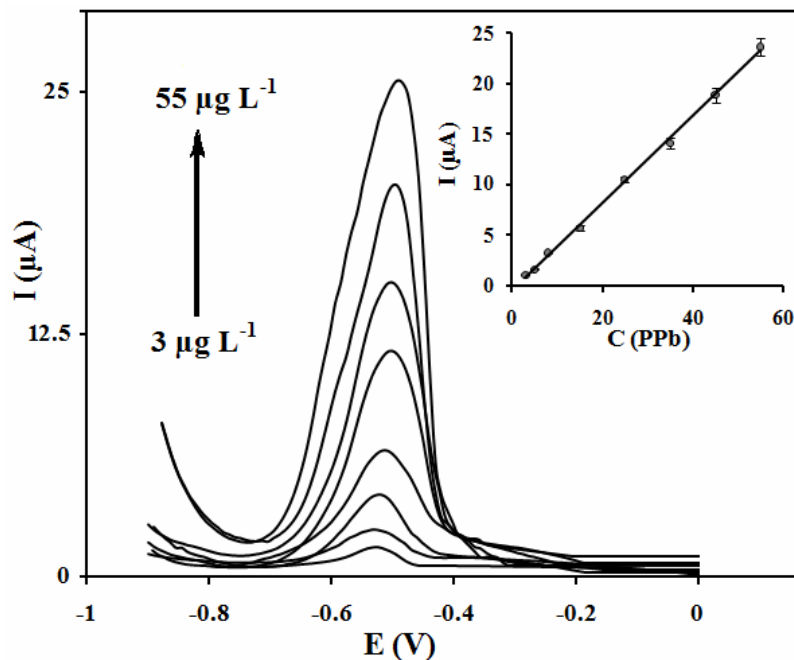


Fig. 9. Stripping voltammograms of the proposed MCPE under optimized experimental conditions, lead(II) concentration ($\mu\text{g l}^{-1}$) 3, 5, 8, 15, 25, 35, 45 and 55. Inset shows the related calibration graph.

Table 1. Effects of Various Metal Ions on the Electrochemical Signals of Lead(II)

Interference	Contribution (%) ^a
Cd(II)	-3.1
Cu(II)	-4.9
Zn(II)	-2.1
Cr(III)	-3.7
Ag(I)	-1.2
Hg(II)	-4.0
Al(III)	-0.8
Fe(III)	-1.7
Mn(II)	-1.2
Co(II)	-0.9
Ni(II)	-2.1

^a $I_{p,Pb(II)} = 100\%$.

Table 2. Comparison of the Performance of the Proposed Lead Sensor with other Electrochemical Sensors

Electrode type	Method	LOD ($\mu\text{g l}^{-1}$)	Linear range ($\mu\text{g l}^{-1}$)	[Ref.]
Cyclodextrin-modified gold electrode	DPSV ^a	1.9	4.6-251	[6]
Glassy carbon electrode modified with MWCNTs/Nafion composite	ASV ^b	1.3	21.6-1621	[28]
Glassy carbon electrode modified with mercury-film	ASV	0.78	5.4-27	[29]
Carbon paste electrode modified with MWCNTs	PSA ^c	6.6	58.4-646.2	[30]
Bismuth modified gold microelectrode	SWASV ^d	3.3	10.8-1810.3	[31]
CPE modified with IIP-MWCNTs	DPSV	0.5	3-55	This work

^aDifferential pulse stripping voltammetry (DPSV). ^bAnodic stripping voltammetry (ASV). ^cPotentiometric stripping analysis (PSA). ^dSquare wave anodic stripping voltammetry (SWASV).

Table 3. Determination of Pb(II) in Different Water Samples

Sample	Added ($\mu\text{g l}^{-1}$)	Found by voltammetry ($\mu\text{g l}^{-1}$)	Recovery (%) ^a \pm R.S.D. ^b	Found by GF-AAS ($\mu\text{g l}^{-1}$)
River water	-	4.2	-	4.4
	6.00	9.9	97.0 \pm 1.9	-
	10.00	14.5	102.1 \pm 2.7	-
Aqueduct water	-	B.D.L. ^c	-	B.D.L.
	6.00	5.8	96.6 \pm 2.5	-
	10.00	9.9	99.0 \pm 2.0	-
Copper factory waste water	-	12.8	-	12.5
	10.00	23.2	98.2 \pm 3.1	-
Coal Processing waste water	-	9.7	-	10.1
	10.00	20.3	103.0 \pm 3.2	-

^aAverage value from three individual experiments. ^bRelative standard deviations (n = 3). ^cB.D.L.: Below detection limit.

Table 4. Analysis of Lead in Synthetic Samples

Sample	Composition	Found ^a	Recovery
Synthetic sample1	Co(II):50.0, Cd(II):50.0, Zn(II):50.0, Al(III):50.0, Pb(II):15.0 $\mu\text{g l}^{-1}$	14.6 \pm 0.4 $\mu\text{g l}^{-1}$	97.3
Synthetic sample2	Cr(III):50.0, Ni(II):50.0, Fe(III):50.0, Cu(II):50.0, Pb(II):15.0 $\mu\text{g l}^{-1}$	14.4 \pm 0.6 $\mu\text{g l}^{-1}$	96.0

^aAverage of three replicate measurements \pm standard deviation.

The effect of the reduction time on the sensor performance was studied. Based on the results of this study, the peak currents increased linearly versus the accumulation time between 10 and 30 s and then leveled off after 40 s. Hence, reduction time of 40 s was applied as an optimum applying time in the present study.

Study of Interferences

Some cations species were examined concerning their interference with the determination of Pb(II) under the optimum conditions. The effect of 200 $\mu\text{g l}^{-1}$ Cd(II), Cu(II), Zn(II), Cr(III), Ag(I), Hg(II), Al(III), Fe(III), Mn(II), Co(II) and Ni(II) on the voltametric response of 10 $\mu\text{g l}^{-1}$ Pb(II) was investigated. As shown in Table 1, most of these metal ions do not interfere with the detection of lead (below 5%). Accordingly, the method has good tolerance towards matrix interferences without sacrificing the signal intensity.

Calibration Curve, Detection Limit and Reproducibility

Figure 9 shows the voltammograms of the MCPE toward Pb(II) with the corresponding calibration plot (inset). The voltammetric peak currents varied linearly with Pb(II) concentration in the range of 3-55 $\mu\text{g l}^{-1}$ with a correlation coefficient, R^2 , of 0.9987. The limit of detection (LOD) was found to be 3 $\mu\text{g l}^{-1}$ based on the signal-to-noise characteristics of $S/N = 3$ (where the S and N are the electrochemical signals in the presence and absence of analyte, respectively). The relative standard deviation (R.S.D., for 10 measuring cycles) was 3.1% demonstrating the excellent repeatability of the proposed method. Another important feature is the functional stability of the

electrochemical sensor. The MCPE, used in this work, were stable during at least one month of intensive use. The elaborated procedure was also compared with those of the some previously analytical methods [6,28-31] for analysis of lead (Table 2). It can be seen that the suggested protocol can provide better sensitivity over the most of the reported methods.

Practical Application

To test the suggested methodology, various real water samples including river water, aqueduct water and waste water were assayed after pre-treatment according to section 2.4. The content of lead(II) was then determined by the standard addition method (Table 3). The recoveries of spiked sample were evaluated using the t-test, and at 95% confidence level. The analytical data were verified by graphite furnace atomic absorption spectrometry (GF-AAS) as a confirmatory method. The reliability of the the proposed procedure was also investigated by the analysis of lead in two synthetic samples (Table 4). Evidently, the electrochemical detection assay can be used for practical analysis of Pb(II) in real samples.

CONCLUSIONS

A novel procedure for selective and sensitive determination of lead(II) ions in the environmental water samples was successfully developed. The presented method possesses some advantages such as simplicity, remarkable selectivity, high sensitivity, cost-effectiveness and good reproducibility. The obtained LOD is lower than the maximum contaminant level of lead allowed by the EPA

(15 $\mu\text{g l}^{-1}$) for drinking water. In light of these findings, the novel electrochemical sensor based on lead IIP material would likely be suitable for application in environmental monitoring. To conclude, the MCPE based on imprinted strategy is very promising.

REFERENCES

- [1] M. Fayazi, M.A. Taher, D. Afzali, A. Mostafavi, M. Ghanei-Motlagh, *Mater. Sci. Eng., C* 60 (2016) 365.
- [2] S. Gunduz, S. Akman, *Food Chem.* 141 (2013) 2634.
- [3] A. Islam, H. Ahmad, N. Zaidi, S. Kumar, *ACS Appl. Mater. Interfaces* 6 (2014) 13257.
- [4] M. Ghanei-Motlagh, M. Fayazi, M. Taher, E. Darezereshki, E. Jamalizadeh, R. Fayazi, *RSC Adv.*, 5 (2015) 100039.
- [5] S.-S. Li, W.-J. Li, T.-J. Jiang, Z.-G. Liu, X. Chen, H.-P. Cong, J.-H. Liu, Y.-Y. Huang, L.-N. Li, X.-J. Huang, *Anal. Chem.* 88 (2015) 906.
- [6] W. Li, G. Jin, H. Chen, J. Kong, *Talanta* 78 (2009) 717.
- [7] A. Shirzadmehr, M. Rezaei, H. Bagheri, H. Khoshsafar, *Int. J. Environ. Anal. Chem.* 96 (2016) 929.
- [8] M. Nasiri-Majd, M.A. Taher, H. Fazelirad, *Talanta* 144 (2015) 204.
- [9] M. Ghanei-Motlagh, M.A. Taher, A. Heydari, R. Ghanei-Motlagh, V.K. Gupta, *Mater. Sci. Eng., C* 63 (2016) 367.
- [10] H.-T. Fan, J. Li, Z.-C. Li, T. Sun, *Appl. Surf. Sci.* 258 (2012) 3815.
- [11] H. Ebrahimzadeh, E. Moazzen, M.M. Amini, O. Sadeghi, *Chem. Eng. J.* 215-216 (2013) 315.
- [12] S. Wei, Y. Liu, M. Shao, L. Liu, H. Wang, Y. Liu, *RSC Adv.* 4 (2014) 29715.
- [13] M. Li, C. Feng, M. Li, Q. Zeng, Q. Gan, H. Yang, *Appl. Surf. Sci.* 332 (2015) 463.
- [14] Q. Li, H. Su, T. Tan, *Biochem. Eng. J.* 38 (2008) 212.
- [15] R. Zhiani, M. Ghanei-Motlagh, I. Razavipanah, J. *Mol. Liq.* 219 (2016) 554.
- [16] M. Shamsipur, H.R. Rajabi, S.M. Pourmortazavi, M. Roushani, *Spectrochim. Acta, Part A* 117 (2014) 24.
- [17] M.K. Bojdi, M. Behbahani, A. Sahragard, B.G. Amin, A. Fakhari, A. Bagheri, *Electrochim. Acta* 149 (2014) 108.
- [18] H. Ashkenani, M.A. Taher, *J. Electroanal. Chem.* 683 (2012) 80.
- [19] A. Bahrami, A. Besharati-Seidani, A. Abbaspour, M. Shamsipur, *Mater. Sci. Eng., C* 48 (2015) 205.
- [20] M. Koneracká, P. Kopčanský, M. Antalík, M. Timko, C. Ramchand, D. Lobo, R. Mehta, R. Upadhyay, *J. Magn. Magn. Mater.* 201 (1999) 427.
- [21] H. Shaikh, N. Memon, M.I. Bhangar, S.M. Nizamani, A. Denizli, *J. Chromatogr. A* 1337 (2014) 179.
- [22] Z.-C. Li, H.-T. Fan, Y. Zhang, M.-X. Chen, Z.-Y. Yu, X.-Q. Cao, T. Sun, *Chem. Eng. J.* 171 (2011) 703.
- [23] H.-T. Fan, T. Sun, H.-B. Xu, Y.-J. Yang, Q. Tang, Y. Sun, *Desalination* 278 (2011) 238.
- [24] Z. Long, W. Xu, Y. Lu, H. Qiu, *J. Chromatogr. B* 1029-1030 (2016) 230.
- [25] T. Alizadeh, *Biosens. Bioelectron.* 61 (2014) 532.
- [26] W. Yang, L. Liu, W. Zhou, W. Xu, Z. Zhou, W. Huang, *Appl. Surf. Sci.* 258 (2012) 6583.
- [27] M. Fayazi, M.A. Taher, D. Afzali, A. Mostafavi, *Mater. Sci. Semicond. Process.* 40 (2015) 501.
- [28] D. Sun, Z. Sun, *J. Appl. Electrochem.* 38 (2008) 1223.
- [29] L.C. Martiniano, V.R. Abrantes, S.Y. Neto, E.P. Marques, T.C. Fonseca, L.L. Paim, A.G. Souza, N.R. Stradiotto, R.Q. Aucélio, G.H. Cavalcante, *Fuel* 103 (2013) 1164.
- [30] C.R.T. Tarley, V.S. Santos, B.E.L. Baêta, A.C. Pereira, L.T. Kubota, *J. Hazard. Mater.* 169 (2009) 256.
- [31] M. Oliveira Salles, A.P. Ruas de Souza, J. Naozuka, P.V. de Oliveira, M. Bertotti, *Electroanalysis* 21 (2009) 1439.